



International Journal of Life Sciences Biotechnology and Pharma Research





Research Paper

ALCOHOL INDUCED CHANGES OF RAT SERUM NITRITE AND NITRATE LEVELS

M Yugandhar^{1*} and K Vijaya Kumari¹*Corresponding Author: **M Yugandhar** ✉ dryugandar.sv@gmail.com

Nitric Oxide (NO) has been named as molecule of the millennium, Number of authors reported alcohol as to impair the normal functions of cardio-vascular NO pathway in experimental model. We report here the impact of 5 g/kg wt of (20%) alcohol on the rat serum nitrite and nitrate s ($\text{NO}_2^-/\text{NO}_3^-$) levels in vivo. Alcohol at the doses administered enhanced rat serum $\text{NO}_2^-/\text{NO}_3^-$ levels continues that this ascent by way of producing free radical type damage may cause myocardial toxicity in rats.

Keywords: Alcohol, Nitrite, Nitrate, Rat Serum

INTRODUCTION

Forgoing provides in formation regarding the production of $\text{NO}_2^-/\text{NO}_3^-$ from the NO and its reactions with other molecules in general and under alcoholic stress. In view an attempt was made to determine the serum $\text{NO}_2^-/\text{NO}_3^-$ levels of both control and alcohol treated rat serum in vivo and the study of these two parameters has been reported to be one of the ways for assessing the overall NO pathway (Guarner, 1993). The data in Table 1 shows that the rat serum receives 5 g/kg of 20% alcohol exhibit an increased levels of both their $\text{NO}_2^-/\text{NO}_3^-$ levels in a statistically significant ($p < 0.001$) manner compared to the normal control group rat serum and the percent increase was found to be more in the rats serum treated with alcohol over 10 weeks period, continuous that the effect of alcohol of rat serum

$\text{NO}_2^-/\text{NO}_3^-$ levels be in a dose and time dependent manner in the control rat serum NO_2^- levels were found to be more than NO_3^- levels and the present observed trend was in accordance with the reports of (Uzun *et al.*, 2005; Yugandhar *et al.*, 2011).

MATERIALS AND METHODS

Albino rats of the weight range 125 ± 5 g were selected for the current study. Animals were maintained at a constant temperature of $15 \pm 5^\circ\text{C}$. The rats were divided into three groups of seven each and were maintained in separate cages. They were fed *ad libitum* with commercial rat feed supplied by Kamadhenu Agencies, Bangalore, India.

Treatment of Animals

For *in vitro* studies 20% alcohol 100-1000 μl was

¹ Department of Zoology, S V University, Tirupati - 517502, AP.

Table 1: Levels of Nitrite and Nitrate in the Serum of Alcohol Treated Rats (Values Expressed as NG NO₂⁻ ml of Serum)

Rat Serum	Control	Alcohol Treated	
		5 Weeks	10 Weeks
Nitrite (NO₂⁻) Levels			
AV	62.336	71.323	87.287
SD	1.406	1.949	3.373
PC		14.417*	40.028*
t-test			
Nitrate(NO₃⁻) Levels			
AV	50.787	68.019	74.270
SD	2.813	2.590	2.806
PC		33.929	46.241
t-test (NO ₃ ⁻)			

Note: Values expressed as the mean ± S.D of 6 samples; AV: Average; SD: Standard deviation; PC: Percent change over control; NS: Not Significant; * p< 0.001.

used. For *in vivo* studies group II rats were gavaged with 5 g/kg body of alcohol 20% (w/v) over 5 weeks (weekly doses). Group III rats were gavaged with 5 g/kg body weight of alcohol over 10 weeks (weekly doses) and group I animals acted as normal control ones.

Determination of Serum Nitrite and Nitrite Levels

The serum was collected by centrifuging the blood at 2000 rpm for 10 min. NO₂⁻ and NO₃⁻ levels determinants were done according to the procedure of Garner *et al.* (1993). Briefly the control and experimental animal's serum samples were deproteinized before analysis with 35% sulfosalicylic acid. Supernatants of serum were analyzed for NO₂⁻ concentration, but determination of NO₃⁻ necessitated reduction of NO₃ to NO₂ which was achieved by reducing the samples in the presence of nitrate reductase and

NADPH. The concentration of NO₂ was determined with the Griess reaction. The Griess reagent consisted of one part 0.1% naphthylamine dihydrochloride and on part of 0.1% sulfanilamide (sulfanilic acid) in 5% phosphoric acid which were mixed and kept chilled. The color was developed by incubation for 1 h at a 60°C and the absorbance was measured in a spectrophotometer at 546 nm. Total nitrite and nitrate concentrations were calculated based on the curve obtained with sodium nitrite standards. NO₃⁻ was determined by subtracting NO₂⁻ values from NO₂⁻ ± NO₃⁻ values (measured as NO₂⁻). The data was expressed as ng of NO₂⁻ /ml of serum.

Statistical Analysis

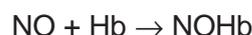
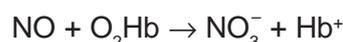
For each parameter, the mean of individual observation (for both control and experimental groups were taken into consideration). Statistical

significance of the data was analyzed through two was ANOVA (analysis of Variance); SNK (Student – Newman - Keuls) test and regression analysis.

RESULTS

The data presented in Table 1 depicts that alcohol at in vivo doses (5 g/kg wt) employed in the present investigation has enhanced both the $\text{NO}_2^-/\text{NO}_3^-$ levels of rat serum. In the control alcohol treated rat serum NO_2^- levels appeared to be more than the NO_3^- levels (Table 1).

The overall pathway of $\text{NO}_2^-/\text{NO}_3^-$ formation is as shown below:



Since $\text{NO}_2^-/\text{NO}_3^-$ are being the end products and part and parts of NO pathway, the author attempted to study the in vivo effect of alcohol on rat serum $\text{NO}_2^-/\text{NO}_3^-$ levels in vivo.

DISCUSSION

Kroncke *et al.* (1997) have reported several cytotoxic effects of NO. NO may react with proteins and nucleic acids. NO has been shown to change ion currents through the mitochondrial membrane leading to release of Ca^{2+} into the cytosol (Nishikawa *et al.*, 1996; Richter *et al.*; 1994). In view of the above mentioned cytotoxic effects of NO end products $\text{NO}_3^-/\text{NO}_2^-$, excess of $\text{NO}_2^-/\text{NO}_3^-$ formation induced by alcohol in the present study (Table 1) may cause any of the

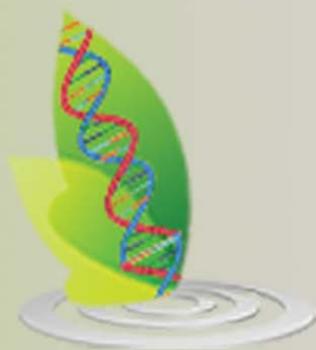
above stated cytotoxic effects in the myocardial system of rat. Oxidative damage correlates with the amount of ethanol consumed (Clot *et al.*, 1994). Recently, it has been demonstrated that NO is an important mediator of hepatotoxicity, and the changes in its generation or actions may contribute to pathologic states (Gardner *et al.*, 1998. Diez - Fernandez *et al.*, 1998). However, in some models of inflammation including alcoholism, it has been shown that impairment of NO due to alcohol stress increases tissue dysfunction or injury (Kubes *et al.*, 2000) due to formation of $\text{NO}_2^-/\text{NO}_3^-$ in the serum.

In the present investigation, elevated levels of alcohol treated serum $\text{NO}_2^-/\text{NO}_3^-$ though difficult to explain the reasons, from the available literature this trend could be due to degradation of organ/cellular based NO in the rats under alcoholic stress. Identical reports were also made by various authors for agents like cyclosporine (Rajeswara Rao *et al.*, 1998), for CCl_4 by Harish Babu (1999); Yugandhar (2011), for organic insecticides by Rajeswara Rao (1997) and for heavy metals by Neelakantam (2008). The excess formation of $\text{NO}_2^-/\text{NO}_3^-$ in the alcohol treated rat serum may cause free radical type stress in alcohol treated rat tissues in part may lead to myocardial toxicity in rats.

REFERENCES

1. Augustinsson K B (1957), *Methods of biochemical analysis*, Vol. 5, Glick D (Ed.), Inter Science Publishers Inc. New York, USA. p. 1.
2. Clot P, Tabone M, Arico S and Albano E (1994), "Monitoring oxidative damage in patients with liver cirrhosis and different daily

- alcohol intake”, *GUI.*, Vol. 35, pp. 1637-1643.
3. Diez-Fernandez C, Sanz N, Alvarez A M, Zaragoza A and Cascales M (1998), “Influence of aminoaguanide on parameters liver injury and regeneration induced in rats by a necrogenic dose of thioacetamide”, *Br J Pharmacol.*, Vol. 125, pp. 102-108.
 4. Gardner C R, Heck D E, Yang C S, Thomas P E, Zhang X J, DeGoerge G L, Laskin J D and Laskin D L (1998), “Role of nitric oxide in acetaminophen-induced hepatotoxicity in rat”, *Hepatology*, Vol. 27, pp. 748-754.
 5. Guarner C, Sariano G, Tomas A, Bulbena O, Nobella M T, Balanzo J, Vilardel F, Maurella M and Monacada S (1993), “Increased serum nitrite and nitrate levels in patients with cirrhosis; relationship to endotoxemia”, *Hepatology*, Vol. 18, pp. 1139-1143.
 6. Harish Babu K (2011), “Reversal of CCl₄ induced toxicity in rat liver by alpha tocopherol”, Ph.D. Dissertation, S.V. University, Tirupati.
 7. Kroncke K D, Fehsel K and Bachofen V K (1997), “Nitric Oxide: Cytotoxicity versus cytoprotection – How, Why, When and Where?”, *Nitric Oxide: Biology and Chemistry*, Apr. Vol. 2, pp. 107-120.
 8. Kubes P and McCafferty D M (2000), “Nitric Oxide and intestinal inflammation”, *Am J Med.*, Vol. 109, pp. 150-158.
 9. Neelekantam C (2007), “Impact of selected heavy metals on the nitric oxide pathway in rats”, Ph.D. thesis, S.V. University, Tiruapti.
 10. Nishikawa M, Sato E F, Utsumi K and Inoue M (1996), *Cancer Res.*, Vol. 56, pp. 4535-4540.
 11. Rao M R, Hutcheson A E and Markov A K (1998), “Changes in the cardiovascular nitric oxide pathway in cyclosporine – A treated rats”, *Drug and Chemi. Toxicol.*, Vol. 21(1), pp. 27-34.
 12. Rao M R, Olinde K D and Markov A K (1998), “In vitro induction of nitric oxide by fructose -1,6 – diphosphate in the cardiovascular system of rats”, *Mol.Cell. Biochem.*, Vol. 185, pp. 171-175.
 13. Richter C, Gogvadze V, Schlaphach R, Schweizer M and Schlegel J (1994), *Biochem. Biophys. Res. Commun.*, Vol. 205, pp. 114-1150.
 14. Uzun H, Zengin K, Taskin M, Aydin S, Simsek G and Dariyerli N (2004), “Changes in leptin. plasminogen activator factor and oxidative stress in morbidly obese patients following open and laparoscopic Swedish adjustable gastric banding”, *Obes Surg.*, Vol. 14, pp. 659-665.
 15. Yugandhar M (2011), “Impact of alcohol on the cardiovascular nitric oxide pathway in rats”, Ph.D. Thesis, Sri Venkateswara University, Tiruap.



International Journal of Life Sciences Biotechnology and Pharma Research

Hyderabad, INDIA. Ph: +91-09441351700, 09059645577

E-mail: editorijlbpr@gmail.com or editor@ijlbpr.com

Website: www.ijlbpr.com

